# SOME EUCOSMINI (TORTRICIDAE) ASSOCIATED WITH EUCOSMA EMACIATANA (WALSINGHAM) and Eucosma Totand kearfott; Four new species, a New combination, and a Neiv SYNONYMY 

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#### Abstract

Eucosma emaciatana (TValsingham) is transferred to Pelochrista Lederer, and Pelochrista perpropinqua (Heinrich) is recognized as a junior synonym of P. emaciatana. Three species considered by previous authors to be superficially similar to emaciatana are reviewed: Eucosma larana (WValsingham), Eucosma totana Kearfott and Pelochrista popana (Kearfott). Four previously unrecognized species are described: Eucosma piperata, new species, Eucosma nordini, new specics, Eucosma taosana, new species, and Pelochrista powelli, new species. Lectotypes are designated for emaciatana and larana. Adults and genitalia of these species are illustrated, and new distributional records are presented.


Additional key words: Oletreutinae, Pelochrista, Nearctic

The Rocky Mountain and Great Basin regions of western United States are home to many similar looking species of Eucosmini, some of which were named by Walsingham in the last quarter of the nineteenth century. Lacking access to the Walsingham types, early North American tortricid specialists often had difficulty applying those names, and in some instances the confusion that resulted has persisted to the present day. One such case involves Eucosma emaciatana (Walsingham), Eucosma larana (Walsingham), and Pelochrista perpropinqua (Heinrich). Heinrich (1923) confused emaciatana with larana (see discussion below) and later (1929) described pcrpropinqua based on a series of specimens of emaciatana. I confirmed that emaciatana and perpropinqua refer to a single taxon by examining the types. Based on male genitalia, the appropriate generic assignment for this species is Pelochrista Lederer, a conclusion reached by Powell (1983) in his placement of perpropinqua, so 1 propose to resolve this situation by transferring cmaciatana to Pelochrista and treating perpropinqua as a junior synonym.

In examining specimens from various institutional and private collections I encountered two previously unrecognized species of Eucosma Hübner that have been confused with larana. They are described below as E. piperata, new species, and E. nordini, new species. Also included are reviews of E. totana Kearfott and $P$. popana (Kearfott), two species considered by previous
authors to be similar in appearance to larana and/or emaciatana. Finally, descriptions are provided for two additional new taxa, Eucosma taosana, new species, and Pelochrista powelli, new species. The former has previously been misidentified as totana; the latter is superficially similar to totana and taosana.
Valsingham (1884) described Paedisca emaciatana from three male specimens collected by H. K. Morrison in Arizona. Fernald [1903] placed this species in Eucosma, and there it has resided ever since. The Fernald collection, acquired by the United States National Museum (USNM) in 1924-25, included two male specimens determined by Walsingham as emaciatana. Neither has an abdomen. One was collected by Morrison in Arizona in 1883 and agrees with the description of emaciatana, the other is lacking collection data and is in such poor condition that I cannot confirm the accuracy of its determination. Heinrich's review (1923) of cmaciatana makes no mention of these two specimens, so I assume he did not examine them. His treatment was based on a series of specimens from Utah, and he illustrated the genitalia (Fig. 193) of a male collected by Tom Spalding at Eureka, Utah, on 27 July 1911. I examined that specimen and a number of other USNM specimens determined by Heinrich as emaciatana and concluded (see discussion below) that they represent E. larana. This explains why Heinrich, when presented with specimens of emaciatana collected in Arizona by O. C.


Figs. 1-12. 1, P. emaciatana, lectotype male. 2, E. larana, lectotype male, 3, E. larana, male, Albany Co., Wyoming. 4, E. piperata, male, Oneida Co., Idaho. 5, E. mordimi, holotype male. 6, E. larana, female, Oneida Co., Idaho. 7, E. totana, male, Grand Co., Colorado. S, E. totana, male, Oneida Co., Idaho. 9, E. taosana, holotype male. 10, P. emaciatana, male, Cochise Co., Arizona. 11, P. popana, male, Larimer Co., Colorado. 12, P. pouelli, holotype male.

Poling, interpreted them as representing a new species, which he described (1929) as E. perpropinqua.

Walsingham (1579) described E. larana from thrce specimens (2 6,19 ) collected in Siskiyou County, California. The forewing (Fig. 2) of the lectotype (designated below) is white with pale brownish-orange markings; that of the female paralectotype is white with a few black specks and only a hint of brownish-orange markings. The specimens misidentified by Heinrich as emaciatana have pale yellowish-white forewings that are generously overlaid with pale brownish-orange coloration. They also have brownish-gray markings (as in Fig. 3). Specimens I collected in southeastern Idaho (Fig. 6) have very pale yellowish-white forewings with just a trace of brownish-orange coloration. I found no substantial differences in the male and female genitalia of these various specimens, and lacking any distinguishing biological information, I concluded that they all represent a single variable species. Curiously, Heinrich (1923) did correctly identify the male that he illustrated (Fig. 197) as larana. Two of the new species described below are similar to larana in forewing color, and each is sympatric with larana in at least a portion of the latter species' range: piperata in Utah and southeastern Idaho, nordini in southeastern Wyoming.

Kearfott (1907) reported a type series for totana consisting of five specimens collected by Tom Spalding and O. C. Poling in Stockton, Utah, and South Utah, respectively. The American Museum of Natural History (AMNH) has a male collected in So. Utah labeled LECTOTYPE, a designation Klots (1942) attributed to Heinrich (1923). I assume from Kearfott's remarks that the collector was Poling, but there is no such indication on the pin labels. I located four Spalding specimens from Stockton, Utah, that are likely to be the other syntypes. Only one is actually totana, a male in the USNM with no capture date. Both it and the lectotype bear the handwritten label "Eucosma totana Cotype Kearf." and Kearfott's printed red label "TYPE Collection of IV. D. Kearfott". A female in the AMNH collected VIII-4-4 and bearing the red Kearfott "TYPE" label is no doubt the specimen referred to by Klots (1942) as a paralectotype, but its genitalia indicates it is not totana. The remaining two, a USNM specimen dated V'III-30-4 and an AMNH specimen dated VIII-14, are conspecific with a series of USNM specimens determined by Heinrich as totana but bearing a hand written label with the notation "dark var." They agree with specimens I collected in New Mexico that are sufficiently distinct in maculation and genitalic details to justify separate species status. Although I was unable to locate females of this taxon, the likelihood of it being confused with totana prompted me to describe it here
as E. taosana. This investigation also brought to my attention the previously unrecognized P. powelli, which can be confused with totana and taosana.

Finally, Kearfott (1907) based his description of popana on 27 syntypes collected by Tom Spalding at Stockton, Utah, with capture dates between 1 June and S August. Klots (1942) reported thirteen specimens in the AMNH as belonging to the type series, including one labeled LECTOTYPE. The lectotype designation should be creditcd to Klots (1942), even though he attributes it to Heinrich (1923). I examined this material and found the lectotype to be a female, rather than a male as stated by Klots. I believe nine of the other twelve specimens are popana, but one has a capture date of 9 August, which is inconsistent with Kearfott's remarks. Of the remaining three, two are males of Epiblema sosana (Kearfott), and one is a female of uncertain identity. Listed below as paralectotypes are the eight AMNH popana specimens whose capture dates agree with Kearfott's statements and seven USNM specimens that I judge to belong to the popana type series.

I am designating lectotypes for emaciatana and larana and have attached designation labels to those specimens. The specimens chosen for this purpose were originally selected by Obraztsov, but his designations were never published.

## Materials and Methods

This study is based on 607 adult specimens and 111 associated genitalia preparations. I examined the types of the five previously described species. Material was borrowed from the following institutional and private collections: AMNH, George J. Balogh (GJB), Canadian National Collection (CNC), Colorado State University (CSU), Essig Museum of Entomology (ENIE), Clifford D. Ferris (CDF), Los Angeles County Museum of Natural History (LACM), Museum of Comparative Zoology (MCZ), John S. Nordin (JSN), The Natural History Museum, London (BMNH), USNM, Donald I. Wright (DJVV), and University of Wroming (UWT). The line drawings were based on images generated by a Ken-A-Tision microprojector (Model X1000-1), and each associated scale bar represents 0.5 mm . Images of the genitalia of the lectotypes of larana and emaciatana were obtained by scanning negatives of photographs taken by Obraztsov of slides he had prepared. All measurements were estimated to the nearest tenth of a millimeter with the aid of a reticule mounted in a Leica MZ95 stereomicroscope. Ratios of measurements were rounded to two decimal places. Forewing length (FIVL) indicates the distance from base to apex, including fringe. Aspect ratio (AR) refers to the ratio of FWL to
foresing width, the latter quantity being measured midway between base and aper. In males, the ratio of forewing costal fold Icngth to FWL is denoted by CFR (costal fold ratio), and the ratio of valval neck width to width of lasal portion of valva by NR (neck ratio). Reported values of AR, CFR and NR are averages of such values calculated for a small sample of specimens. The number of items supporting a particular statistic is indicated by $n$.
Some species discussed here only vaguely display the putative, ancestral, fasciate forewing pattern for the Tortricidae discussed by Brown and Powell (1991) and Baixeras (2002), but their terminology is used when possible in the forewing descriptions.

## SPECIES ACCOUNTS

## Eucosma larana (Walsingham)

(Figs. 2, 3, 6, 14, 15, 25, 36)
Paedisca larana Walsingham 1879:43.
Eucosma larana: Fernald [1903]:456; Barnes and McDunnough 1917:169; Heinrich 1923:110; McDunnough 1939:47; Powell 1983:34.
Eucosma emaciatana: (not Walsingham 1884) Heinrich 1923:108; McDunnough 1939:46; Powell 1983:34.
Types. Lectotype here designated (Figs. 2, 14): \%, Sheep Rock, Siskiyou Co., California, Walsingham, 3 Sept. 1871, genitalia slide 11502, BMNH. Paralectotypes: same data as lectotype ( $1 \mathrm{o}, 1$, O , genitalia slide 11759 , BMNH).
Diagnosis. Darkly marked specimens of larana can be recognized by forewing pattern (Fig. 3), but pale specimens (Fig. 6) might be confused with nordini or piperata. The forewing of nordini (Fig. 5) is pale yellowish white, has a gray streak on the costal fold, and shows no indication of brownish-orange mottling. The combination of white forewing color, black speckling, and brown costal marks distinguishes piperata (Fig. 4). One can also separate larana, piperata, and nordini by the shapes of the sterigmata (Figs. 36, 35, 31) and by
subtle but consistent differences in valval shape (Figs. $15,18,21$ ).
Description. Itead: Yery pale yellowish white, long scales of upper frons and vertex often witl pale gray shading preceding whiter apices; labial palpus with pale-gray lateral surface, third segment enclosed by long narrow seales of second segnent; antema yellowish white. Thoras: Dorsal and ventral surfaces coneolorous with head, legs yellowish white to pale yellowish brown. Forewing (Figs. 2, 3, 6): ${ }^{3}$ FWL $5.5-12 \mathrm{~mm}$ (mean $=10.5, \mathrm{n}=13$ ), $\mathrm{AR}=3.16, \mathrm{CFR}=0.3,8$ FWL S.5-11.1 mm (mean $=10, \mathrm{n}=10$ ), $\mathrm{AR}=3.05$; eosta nearly straight, apex mildly acute, termen straight; dorsal surface white to pale yellowish white and variably overlaid with pale brownisl-orange motting, darker speeimens witl four lrownish-gray marks, the first a triangular pretomal mark on dorsum, often with a few black scales at its anterior extremity, the second a subbasal mark on fold that is ussually comected to dorsum by brownish-orange scaling, the third at distal end of cell, often divided longitudinally by a brownisl-orange streak, the fourth anterior to ocellus; ocellus obscure, variably overlaid with Brownish-orange scales and crossed longitudinally by ip to four black dashes; distal one-half of costa with four, obscure, paired, white strigulae; made costal fold usually grayer than adjacent portion of wing, fringe white to yellowish white, with gray shading near apex and brownish-orange suffusion near tornus. Hindwing: Uniformly pale brownish gray with lighter fringe. Male genitalia (Fig. I4, I5): Uncus divided medially into two variably developed setose lobes with eonvex lateral margins; dorsolateral shoulders of tegumen well developed, often rounded and hunched, sometimes with angular comers; soeii long, flat, and densely setose; vesiea with 2-7 deciduous cornuti ( $\mathrm{n}=$ 10); valva with costal margin concave, apex and ventral angle evenly rounded, distal margin very weakly eonvex, ventral invagination moderate, $\mathrm{NR}=0.61$, cucullus of nearly uniform width, medial surfaee densely setose, sacculus sparsely setose margin of basal opening with patch of short slender spines. Female genitalia (Fig. 25): Papillae anales faeing ventrolaterally and densely setose, medial margins very weakly sinuate, surfaces finely ridged transversely, long setae on lateral margins eurving ventrally, remaining setae shorter with hooked apices; tergum VIII sparsely setose; lamella antevaginalis (Fig. 36) ringlike and weakly sclerotized, lamella postvaginalis with semitriangular posterolateral comers; membrane between sterigma and ventral extremities of tergum VIII setose; sternum VII with posterior margin approximate to sterigma and roundly invaginated to depth of one-half length of sterigma; ductus bursae strongly constricted anterior to ostium, widening anteriorly; corpus bursae with large signum near juneture with ductus bursae and small spike-shaped signum on opposite wall.

Distribution and biology. I examined 53 specimens ( 40 है, $13 \%$ ) from the following states and counties: CALIFORNIA: Siskiyou, Tulare; IDAHO: Oneida; UTAH: Juab; WYOMING: Albany. The flight period extends from early July to the begiming of September, and capture sites range in elevation from $5000^{\prime}$ to $8000^{\prime}$.


FIGS. 13-14.Genitalia of leetotypes. 13, P. emaciatana, slide BMNH 11571. 14, E. larana, slide B.MNH 11502.


Figs. 15-23. Male genitalia. 15, E. larana, slide DJV 763. 16, E. totana, slide DJW 1022, 17, P. emaciatana, slide DJW 952. $18, \boldsymbol{E}$. piperata, slide DJW 762. 19, E. taosana, slide DJW 1035. 20, P. popana, slide DJW 1065. 21, E. nordini, slide DJW 760.22, P. pourelli, slide DJW 1027. 23, P. powelli, slides DJW 1032 and 705

This moth has been collected in open sage brush habitat in Idaho and Wyoming. No larval host has been reported.

Comments. The variation in forewing color appears to have a geographic component Specimens from northern California and southeastern Idaho have very pale yellowish-white forewings with pale to nearly obsolescent brownish-orange markings, those from Wyoming tend to be darker, with brownish-gray markings and extensive brownish-orange mottling, and those from Utah and central California appear to be intermediate. The medial division of the uncus varies from an inconspicuous line to the pronounced indentation illustrated in Figure 15.

Eucosma totana Kearfott
(Figs. 7, 8, 16, 28, 32)

Eucosma totana Kearfott 1907:32; Bames and McDunnough 1917:169, Heinrich 1923:10S; McDunnough 1939:46; Powell 1983:34.
Eucosma spodias: Meyrick 1912:35.
Types. Lectotype designated by Heinrich (1923): ${ }^{\circ}$, South Utah, July 1900, genitalia slide CH, 2 Dec 1919, AMNH Paralectotype ó: Stockton, Utah, Tom Spalding, USNM.
Diagnosis. This species can be confused with taosana, popana and powelli, but the following combination of dark brown forewing markings usuallysuffices for diagnosis: a subbasal mark on fold, a thin line along fold from subbasal mark to tornus, a pretornal triangular mark based on fold, and a cherron shaped mark at distal end of cell. Some specimens do not show the linc on the fold. Superficially; totana is most similar to taosana (Fig. 9), but the latter species does not have
a dark line on the fold or a chevron shaped mark at the end of the cell. Male genitalie characters separating totana and taosama include: subtle differences in shape of cucullus (Figs. 16, 19), distinctly different shape of uncus, and number of cornuti in vesica ( 10 for totana vs. 5 for taosana). Eucosma totana is easily separated from popana and powelli by the presenee in the latter two species of a stout spine at the ventral angle of the cucullus. Females of taosana are not known, but the sterigmata (Figs. 32, 37, 34) of totana, popana and

powelli are easily distinguished from one another.
Description. Head: Scales of frons and vertes white, sparsely marked with pale gray; labial palpus porrect length ea $3 \times$ eye diameter, second segment with medial surface and dorsal margin white, lateral surface pale brownish gray, scales of ventral and dorsal margins long and slender, concealing third segment; antenna white, Thorax: Scales of dorsal surface and tegulae white basally and apically, brownish gray medially; producing a speckled effect; ventral surface and hindlegs white, fore and midlegs white posteriorly, brown to pale brown anteriorly, with white annular markings on tarsus and tibia. Forewing (Figs. 7,5 ): ${ }^{2}$ FWL $8.7-12 \mathrm{~mm}($ mean $=10.1, \mathrm{n}=65$ ), $\mathrm{AR}=3.2, \mathrm{CFR}=0.27,9 \mathrm{FWL} 7.4-10.5 \mathrm{~mm}$ (mean $=8.9, \mathrm{n}=18$ ), AR $=3.11$; costa nearly straight, apex acute, termen weakly consex; dorsal



Figs. 26-27. Female genitalia. 26, E. piperata. slide DJVV 1061. 27, E. nordini, slide DJVV 1073
surface white with brown to brownish-black irrorations on basal twothirds and five brownish-black marks, the first a subbasal mark on fold, sometimes extending weakly to dorsum, often bordered distally by a thin line of black scales, the second a very thin line along fold from subbasal mark to tomus, the third a triangular mark based on fold and projecting anteriorly along basal margin of ocellus, the fourth a longitudinally elongate mark anterior to ocellus, narrowing basally and extending into cell, the fifth a chevron on distal margin of cell, sometimes connecting anteriorly to mid costa and posteriorly to line on fold, often divided medially by aforementioned longitudinal mark; ocellus bordered on basal, distal and tornal margins with lustrous, pale, yellow-brown to yellow-gray bars, white central field crossed by up to four, black, longitudinal dashes; costal margin brownish black,
crossed by numerous paired white strigulae, numbers four through nine usually sharply delineated; dorsal margin with $10-12$, small, evenly spaced, brownish-black marks; scales along terminal margin white with subapical black markings, fringe usually whiter with more extensive dark markings between M1 and M3. Abdomen: Scales on posterior margin of eighth segment in females brownish black with white apices. Hindwing: Uniformly pale brownish gray with lighter fringe. Male genitalia (Fig. 16): Uncus triangular, dorsal surface setose, posterior surface developed into medial wedge-shaped ridge, dorsolateral shoulders of tegumen well developed; socii long, flat, and densely setose; aedeagus tapered distally, vesica with ca. 10 deciduous cornuti ( $\mathrm{n}=13$ ); valva with costal margin concave, apex and ventral angle evenly rounded. distal margin convex, invagination of ventral


FIGS. 28-29. Female genitalia. 28, E. totana, slide DJW 1028. 29, P. popana, lectotype, slide DJW 1055.
margin moderate, $N R=0.6$, cucullus witl densely setose medial surface, sacculus and margin of basal opening moderately setose. Female genitalia (Fig. 28): Papillae anales facing laterally and densely setose, surfaces finely ridged transversely, setae on medial inargins with hooked apices, those on lateral margins twice as long and curving ventrally; posterior margin of tergum VIII with $3-4$ rows of setae; lamella antevaginalis (Fig. 32) ringlike and very weakly sclerotized, lamella postvaginalis well developed, width of posterior margin ca. $2 \times$ ostium diameter, posterolateral comers acute, a depressed trough from mid posterior margin to ostium; membrane between sterigma and ventral extremities of tergum VIII setose; stemum VII with length of posterior margin ca. $3 \times$ ostium diameter, roundly invaginated to depth of one-third length of sterigma, approximate to sterigma medially; ductus bursae constricted anterior to ostium; corpus bursae with two signa

Distribution and biology. My study sample included 128 specimens ( $1050^{6}, 23$ ? ) from the following states and counties: ARIZONA: Coconino;

COLORADO: Chaffee, Fremont, El Paso, Grand; IDAHO: Lincoln, Oneida; MONTANA: Jefferson; NEIV MEXICO: Santa Fe; OREGON: Harney; UTAH: Juab, Sanpete; IVYOMING: Albany. I have occasionally found this species to be abundant in sagebrush habitat at elevations between $5000^{\prime}$ and $8000^{\prime}$. Brown et. al. (1983) reported Chrysothamnus nauseosus (Pall.) Britt. (Asteraceae) as a larval host in Idaho.

Comments. The forewing markings are stable, but the overall appearance of totana varies from very pale tan to medium brown. In lighter specimens the brown irrorations are restricted to the basal one-third of the wing, the median area is mostly pale yellowish white to white, and the markings are orangish brown. Darker
specimens are much more densely irrorated and have dark brownish-black markings.

## Eucosma piperata Wright, new species

(Figs. 4, 18, 26, 35)
Diagnosis. Reasonably fresh specimens of this species are readily identified by forewing pattern: white, peppered with minute black specks, with brown marks on distal one-half of costa.

Description. Head: Frons and vertex white, a small patch of palebrown scales anterior to eye; labial palpus with medial surface white, lateral surface pale brown; antenna white, scape sometimes pale brown dorsall: Thorav: Dorsal and ventral surfaces white; legs with anterior surfaces pale brown, posterior surfaces white, and distal ends of tarsal segments lightly ringed with white. Forewing (Fig. 4): of FWL $10.5-11.5 \mathrm{~mm}$ (mean $=11.2, \mathrm{n}=6$ ), $\mathrm{AR}=3.11, \mathrm{CFR}=0.32$, FVL 11.5-13.5 mm (mean $=12.6, \mathrm{n}=\mathrm{S}$ ), $\mathrm{AR}=2.99$; distal threefourths of costa straight, apex mildly acute, termen weakly convex; dorsal surface white, sparsely speckled with black scales between radial vein and dorsum, costal strigulae delimited by narrow, brown, costal marks but otherwise not distinguishable from ground color, a conspicuous, oblique, brown mark at apex; ocellus obscure, variably marked on basal and distal margins by a few black and brown scales, respectively, ca. three, black, weakly expressed. longitudinal dashes in central field; costal fold of male pale grayish brown along costal margin; fringe scales white basally, very pale orange brown distally.
Hindwing: White, a shade grayer than forewing, fringe white. Male genitalia (Fig. 1S): Uncus a semicircular dorsally setose lobe, supported laterally by well developed and mildly hunched shoulders; socii densely setose, tapered distally, with lateral margins variably serrate; gnathos a narrow band; vesica with ca. 14 deciduous comuti ( $\mathrm{n}=4$ ); valva with costal margin concave, apex rounded but moderately acute, distal margin convex, ventral angle gently rounded, ventral invagination shallow, $\mathrm{NR}=0 . \overline{\mathrm{T}}$, cucullus with medial surface densely setose, sacculus sparsely setose, a patch of setae on margin of basal opening. Female genitalia (Fig. 26): Papillae anales facing laterally and densely setose, medial margins weakly sinuate, surfaces finely ridged transversely, setae toward lateral margins strongly curved ventrally, those near anal opening with hooked apices; posterior onehalf of tergum VIII with ca. four rows of setae; lamella antevaginalis (Fig. 35) ringlike; lamella postvaginalis widening posteriorly to ca. $2 \times$ ostium diameter, with triangular, mildly setose, posterolateral comers; sternum VII with posterior margin roundly invaginated to threefourths length of sterigma and approximate to sterigma; ductus bursae weakly constricted anterior to ostium, gradually widening anteriorly; corpus bursae with two similarly shaped signa, membrane finely wrinkled near signa, interior surface of bursa minutely microtricliate.

Holotype. ठै, Vineyard, Utah, 9 July 1912, Tom Spalding, genitalia slide USN゙M 70391, ÚSNM.

Paratypes. ARIZONA: Lupton, A. K. Wyatt, 3 July 1951 (1 9, genitalia slide DJW 1061). CALIFORNIA: Inyo Co., 9 mi. W. Lone Pine, P. D. Hurd \& J. A. Powell, 19 July: 1961 ( 1 ) ; ; Westguard Pass White Mts., P. Opler \& J. A. Powell, I9 July 196S (1 \&). COLORADO Mesa Co., Colo. N. Monument, Head of Red Canyon, J. Moore, 3 July 2001 ( 3 b , 1 \& , $0^{\circ}$ genitalia slide DJW 1132, $\%$ genitalia slide DJV1133). IDAHO: Oneida Co., Curlew NG, 4 mi ENE of Holbrook, 5050', D. J. Wright, 18 July 2001 ( 16 ', genitalia slide DJW 762). NEVADA: Nye, Co., Currant Cr. Cpgd., J. Scott, 20 July 196 S ( 1 \&), P. Opler \& J. A. Powell, 20 July $1965^{\circ}$ (2\%, genitalia slides T. Gilligan 265, DJW I2s8); Lincoln Co., Cathedral Gorge St. Pk., I. Doyen, 12/13 July 1971 (1 \%). OREGON: Baker Co., Burnt River Cyn., 3200', $44 \mathrm{E} 33.08^{\prime}$ N, I17E $39.75^{\prime}$ W, C. D. Ferris ( 1 \& , genitalia slide DJW 1134). UTAH Vineyard, 4 July 1912 (1 \&); Vineyard, Tom Spalding, 14 July 1912 (1 d) genitalia slide DJW 1058); Juab Co., Eureka, Tom Spalding, 20 July
 genitalia slide DJW' 1060). Paratype depositories: CDF, CSU, EME LACM, USNM, DJW

Etymology. The specific epithet, deriving from the Latin word for
pepper, refers to the minute black speckling on an otherwise white forewing.

Distribution and biology. The 20 specimens ( 9 d, 11 f) reported above suggest that the range of this moth may be restricted to the Great Basin. The flight period extends from mid June through July. The larval host is unknown.

Comments. In some specimens the black speckling is barely discernable, but the brown costal marks, particularly the apical onc, are usually conspicuous. The wrinkling of the membranc of the corpus bursae in the vicinity of the signa is variable.

## Eucosma nordini Wright, new species <br> (Figs. 5, 21, 27, 31)

Diagnosis. The immaculate pale yellowish-white forewing is diagnostic for this species. Males have a blackish-gray streak along the anterior edge of the costal fold.

Description. Head: Upper frons and vertex very pale yellorish white, labial palpus white, lateral surface of second segment with pale gray shading; antenna white. Thoras: Dorsal surface pale vellowish white, ventral surface white, legs pale vellowish white, anterior surfaces sometimes darker. Forewing (Fig. 5): © FWL $10-13.7 \mathrm{~mm}$ $($ mean $=11.9, \mathrm{n}=14), \mathrm{AR}=3.35, \mathrm{CFR}=0.29, \& F W \mathrm{~F} 10 . S-13 \mathrm{~mm}$ (mean $=11.5, \mathrm{n}=5$ ), $\mathrm{AR}=3.13$; costa straight, apex acute, terminal margin weakly convex; dorsal surface very pale yellowish white, without markings, fringe white, males with blackish-gray streak along costal edge of fold. Hindwing: Pale brownish gray, fringe white. Male genitalia (Fig. 21): Uncus semitriangular, apex rounded, lateral margins weakly convex, dorsal surface setose, shoulders of tegumen well developed; socii long, flat, tapering distally, and densely setose; gnathos a narrow band; aedeagus tapering distally, vesica with 4-10 deciduous comuti ( $\mathrm{n}=5$ ); valva with costal margin concave except for slight, elongate, convex protrusion on cucullus, apex semirectangular, distal margin convex, ventral angle gently rounded, ventral invagination shallow, $\mathrm{NR}=0.75$, cucullus with medial surface densely setose, sacculus and margin of basal opening moderately setose. Female genitalia (Fig. 27): Papillae anales facing laterally and densely setose, surfaces finely ridged transverselv, medial margins mildly sinuate, long setae on lateral margins curving ventralls; setae near anal opening shorter with hooked apices; three to four rows of setae on posterior one-third of tergum VIII; lamella antevaginalis (Fig. 31) ringlike; lamella postvaginalis with triangular, setose, posterolateral, comers, posterior margin weakly invaginated medially; sternum VIl with posterior margin slightly wider than sterigma: roundly invaginated to one-third length of sterigma and approximate thereto; ductus bursae strongly constricted anterior to ostium, gradually widening toward corpus bursae, compus bursae with large signum near juncture with ductus bursae and sinaller spike-like signum on opposite wall posterior to mid bursa, inner sufface minutely microtrichiate.

Holotype. ठ, Wyoming, Albany Co., Medicine Bow NF, 11.5 mi SE Laramie, Jctn. Forest Rds 707 and 705,4 August 2001, D. J. Wright, S220', genitalia slide DJW 760, deposited in USNM. Type locality at $41^{\circ} 11.75^{\prime} \mathrm{N}, 105^{\circ} 23 . \mathrm{T}^{\prime} \mathrm{W}$

Paratypes. COLORADO: Chaffee Co., Salida, G. M. and J. L. Sperry, 24 August 1938 ( 1 \& , genitalia slide DJW1165). WTOMHEG: Albany Co., T15N S73W Sec. 1, $7450^{\prime}$, C. D. Ferris, 25 July 2000 ( 1 ठ), 28 July 2000 ( 1 d , genitalia slide USNM 91928 ), 30 July 2003 ( $10^{\circ}$ ), 5 August 2003 (2 ठ ) , 9 August 2002 ( 1 ó), 9 August 2003 ( 1 ठ́), 10 August 2002 ( $3 \delta^{\circ}$, genitalia slide DJWi077), I0 August 2003 ( $10^{\circ}$ ), 11 August 2003 ( 1 §), 13 August 2002 ( $1 \delta^{*}$ ), 14 August 2002 ( 1 §*), 15 August 2002 (3 $\delta^{\circ}$ ), 15 August 2003 ( $20^{\circ}$ ), 16 August 2002 ( $50^{\circ}$ ), 17


Angust 2002 ( $5^{2}, 1$ ) , 19 August 2003 ( 1 , genitalias slide DJW 107S), 22 . tugust 1999 (1 \%), 22 August 2002 (1 \% genitalia slide DJW 1073). 24 Augist 2002 ( 1 ), 25 August 2002 (1 , genitalia slide DJW 1074); Albani Co, T15N S73W Sec, 1,2217 Sky View Lan, T465', J. S. Nordin. 27 July 1994 ( 1 . genitalia slicle DJ1゙ 312), 11 August 1995 (1 S. genitalia slide D) 11271 ), 14 August 2001 ( 1 3), 16 August 1999 ( 1
 21 August 1998 ( 1 ó, genitalia slide J. W: Brown 1173), $2 S$ August 1998 (1 ©); Albany Co., T15N S7lW Sec. 14, E of Pilot ILill Road, S600', J. S. Nordin, 25 August 1998 ( $20^{\circ}$ ); Albany Co., Upper Blair PG, N of Rd. 705. J. S. Nordin. S200', 12 August 2003 (1 P); Albany Co., NE of Pole Mtn., S. of Happy Jack Rd., S320', J. S. Nordin, 12 August 2001 ( 1 ó); Albany Co., 1.5 mi NW Woods Landing, Fox Creek, J. S. Nordin. 7600 . 31 July 2002 (1 ふ'); Albany Co., S mi. NE Laramie, Rogers Canyon, M. Pogue, 22 August 1950 (3 P, genitalia slide DJII 1135); Albany Co., Nedicine Bow NF, 10.5 mi SE Laramie, S300', D. J. Wright, 4 August 2001 (20); Albany Co., Medicine Bow NF, 11.5 mi SE Laramie, S220', D. J. Wright, 4 August 2001 (1 ©); Teton Co., Grand Teton NP, Teton Sciences School, P. A. Opler, 3 August 2001 ( 1 ő); Washakie Co., Tensleep Preserve, T47N RS6W S32, 6400', S August 1999, C. D. Ferris (3 ठ) . Paratype depositories: AMNH, BMNH, CNC, CDF, CSU, EME, JSN, LACM, USNM, DJW, UWY.

Etymology: lt is a pleasure to name this species after John S Nordin, whose extensive collecting around Laramie, Wyoming, has made a significant contribution to our knowledge of the lepidopteran fauna of that region.

Distribution and biology. I examined 115 specimens ( 104 of 11 \&) from the following states and counties: COLORADO: Chaffee; WYOMING: Albany, Teton, Washakie. Capture sites range in elevation from $6400^{\prime}$ to $\$ 300^{\prime}$. Flight occurs from late July to the end of August. The larval host is unknown.

## Eucosma taosana Wright, new species

(Figs. 9, 19)
Diagnosis. The forewing of taosama has a conspicuons band of orange-brown scales along the costa and a line of similarly colored scales along $1 \mathrm{~A}+2 \mathrm{~A}$. The male genitalia of taosana is similar to that of totana (Figs. 16, 19), but the apex of the cucullus is more angular and the uncus lacks a wedge shaped posterior projection.

Description. Head: Lower frons white, scales of vertex brownish gray medially, lighter toward base and apex; lalbial palpus with medial surface white, lateral surface brown; antenna brown. Thorax: Dorsal surface brown, scales on apex of tegnlate brownish black with white apices, ventral surface pale tan, legs with anterior surfaces dark graybrown. posterior suffaces pale tan, distal extremities of tarsal segments ringed with pale tan. Forewing (Fig. 9): ठ FWL $7.5-9.5 \mathrm{~mm}$ (mean = $\mathrm{S} .5 .31=12), \mathrm{AR}=3.19, \mathrm{CFR}=0.31$; costa and termen nearly straight, apex acnte; clorsal surface brown with brownish-black markings, a band of orange-brown coloration from base to apex between costa and radial vein, a narrow similarly colored band from base to tormes along $1 A+2 A$, a brownish-black, outwardly oblique, subbasal mark on dorsum extending forward into cell, a triangular, brownish-black, subtornal mark on dorsum projecting anteriorly along basal margin of ocellus, both marks divided by orange-brown line along $1 \mathrm{~A}+2 \mathrm{~A}$, a narrow elongate patch of white-tipped, dark grayish-brown scales anterior to ocellus, extending and tapering basally to middle of cell, mildly constricted at distal end of cell; ocellus with basal, distal and tornal margins pale yellowish brown to yellowish gray, central field white to pale brown, crossed longitudinally by 3-4 brownish-black dashes, the latter often connected in zig-zag pattern; distal one-half of costa usually with four, sharply defined, paired, white strigulae, costal fold on male brownish black; termen with band of white-tipped
brommish-black scales from apex to tomos, fringe scales similarly marked near apex. lighter and more uniforml brownish gray toward tormus. Male genitalia (Fig. 19): Uncus trimgular and dorsally. setose, divided medially by shallow indentation: dorsolateral shoulders of tegumen well developed and hunched: aedeagus long, slender, and tapering distally, sesica with $4-5$ decidnous commti ( $n=9$ ); gnathos a narrow loand; valva with costal margin concave, apex rounded but moderately acute, distal margin convex, with ca. $S$ stont setae along ventral two-thirds, ventral angle rounded, ventral invagination moderate, $\mathrm{NR}=0.61$, cucullus with distal one-half of medial surface densely setose, sacculus moderately setose, margin of basal opening with setose medial projection.

IIolotype. ó, New Mexico, Taos Co., S. Side US 64, 10 mi. SE Tres Piedras, 7550 ', 11 August 1999, D. J. Wright, genitalia slide DJW 1035, deposited in USNM. Tipe locality at $36^{\circ} 34.5^{\prime} \mathrm{N}, 105^{\circ} 45.2^{\prime} \mathrm{W}$.

Paratypes. NEW MEXICO: Same data as holotype ( 17 ó, genitalia slides DJW 52S, 1034); Luma Co., Demíng, 16-23 August (2 genitalia slides USNM 70399, DJW 1070); Fort Wingate, 24-30 June ( 1 . genitalia slide USNM1 70396), 24-31 July ( 30 , genitalia slide USNM 70.39S). UTAI: Tooele Co., Stockton, Tom Spalding, 30 August 1904 ( 1 ó, genitalia slide DJIV 1064). Paratype depositories: AMNH, BMNH, CNC, CSU, EME, LACM, USNM, DJIV.

Etymology. The specific epithet refers to Taos County, New Nexico.

Distribution and biology. Of the 29 specimens cxamined, one is from central Utah and the rest are from New Mexico. Capture dates range from late June through August. The type locality is open sagebrush habitat at an altitude of 7550 feet. The larval host is unknown.

## Pelochrista emaciatana (Walsingham), new combination

(Figs. 1, 10, 13, 17, 24, 33)
Paedisca emaciatana Walsingham 1S84:137, pl. IV, Fig. 7.

Eucosma emaciatana: Fcrnald [1903]:460; Barnes and McDunnough 1917:171; McDunnough 1939:46; Powell 195:3:34.

Encosma perpropinqua: Heinrich 1929:S;

## McDunnough 1939:47, new synonymy. <br> Pelochrista perpropinqua: Powell 1983:35.

Types. Pacdisca emaciatana. Lectotype here designated (Figs. 1, 13): ©, Arizona, Morrison, 1882, genitalia slide 11571 , BMNH. Paralectotypes: same data as lectotype, (2 ©, BMNH). Eucosma perpropinqua. Holotype: ơ, Arizona, Pima Co., Indian Oasis, Sells Post Office, 15-30 April 1923, O. C. Poling, genitalia slide 72797, USNM. Paratypes: same site and collector as holotype, 1-15 April 1923 (39, genitalia slides DJW 809 , 955, USNM; 1 \& CNC), 15-30 April 1923 (1 $\%$, USNM; I ${ }^{9}$, AMNH).

Diagnosis. Reasonably fresh specimens can be identified on the basis of forewing pattem (Fig. 10), but dissection is recommended for positive determination. Males are distinguished by the general shape of the valva and the size and position of the ventral spike (Fig. 17), females by the sclerotized plate on the dorsolateral surface of the corpus bursae, the presence of only one
signum, and the shape of the stcrigma (Figs. 24, 33).
Description. Head: Frons and vertex white, scales antcrior to eye light brown; labial palpus elongate, lateral profile triangular, length more than $2 \times$ eve diameter, first segment and medial surface of second segment white, scales on lateral surface and dorsal margin of second segment pale brown with white apices, scales of second segment concealing third segment; antenna white. Thorax: Dorsal surface white with pale-brown shading, scales of tegulae light brown with white apices, ventral surface white, legs light brown with white tarsal ammulations. Forewing (Figs. 1, 10): 6 FWL 10.8-13 mm (mean $=11.5, \mathrm{n}=7), \mathrm{AR}=3.1, \mathrm{CFR}=0.27,9$ FWL $7.9-9.9 \mathrm{~mm}($ mean $=9.4$, $\mathrm{n}=6$ ), $\mathrm{AR}=3.3$; costa straight, vertex acute; dorsal surface white with brown markings, appearing streaked longitudinally, a dark-brown, outwardly oblique, subbasal mark on fold, a brown pretornal mark on dorsum, often one or more variably expressed, disjunct, brown marks between mid costa and pretomal mark, and an elongate patch of pale brown scales with white apices anterior to ocellus and connected by oblique spur of similar scaling to brown apical mark; ocellus obscure, white to pale brown, crossed longitudinally by three light-brown streaks, the latter often marked medially by a few brownish-black scales; termen with several rows of pale brown scales with white apices, the latter preceded basally by a streak of white scales from distal margin of ocellus to apex; distal one-half of costa with four indistinct, white, paired strigulae, merging into light post-costal streak from mid costa to apical mark; male costal fold darker than adjacent forewing scaling. Hindwing: Uniformly brownish gray with pale white fringe. Male genitalia (Figs. 13, 17): Uncus a dorsally setose convex lobe, shoulders of tegumen moderately developed; socii curving dorsally and moderately setose; aedeagus tapered distally, resica with no indication of comuti ( $\mathrm{n}=10$ ); valva with costal margin concave, apex evenly rounded, outer margin convex, ventral angle with well developed projection supporting a stont spine, usually a spine of similar size and several smaller spines on distal margin of cucullus, ventral invagination moderate, $\mathrm{NR}=0.7$, cucullus with distal two-thirds of medial surface densely setose, sacculus moderately setose. Female genitalia (Fig. 24): Papillae anales facing laterally and densely setose, medial margins mildly sinuate, surfaces finely ridged transversely, long setae on lateral margins strongly curved ventrally; lamella antevaginalis (Fig. 33) ringlike and very weakly sclerotized; lamella postvaginalis well developed, depressed medially, with variably invaginated posterior margin and sharply acute anterolateral projections; stemum VII with posterior margin weakly invaginated and closely approximate to sterigma; ductus bursae uniformly narrow; constricted anterior to ostium; corpus bursae with large sclerotized patch on dorsolateral surface at juncture with ductus bursae, a large signum on ventral surface, and a faint indication of a reduced signum at center of anterior margin of sclerotized patch.

Distribution and biology. I examined 96 specimens ( 66 d., 30 ) : one each from San Bernadino Co., California, Clark Co., Nevada, and Kimble Co., Texas; the rest (to the extent determinable by specimen data) from Cochise, Pima, Pinal, and Santa Cruz Counties in Arizona. Three specimens had capture dates in September or October; the others were collected between early March and mid June. Ninety percent of the records were dated between 1 April and 31 May. No larval host information has been reported.

## Pelochrista popana (Kearfott) <br> (Figs. 11, 20, 29, 37)

Eucosma popana Kearfott 1907:31; Barnes \& McDunnough 1917:169; Heinrich 1923:109; McDunnough 1939:47.

Eucosma carcharias: Meyrick 1912:35.

## Pelochrista popana: Powell 195:3:3.5.

Types. Lectotype designated by Klots (1942): ₹, Stockton, Utah, Tom Spalding, 3 July 1904, genitalia slide DJW 1085, ANNH. Paralectotypes: UTAH: Stockton, Tom Spalding, 1 June 1904 (36, AMNH; 1 d, USNM), 8 June 1904 ( 2 ó, USNM), 10 June 1904 (2 2 , USNM), 14 Junc 1904 (2 2 , AMNH; 1 ठ, USNM), 15 June 1904 ( 1 oै, USNM), 28 June 1904 ( 1 ค, AMNH), 3 July 1904 (2 $\%$, AMNH).
Diagnosis. The forewing maculation of popana is grayish-black, as opposed to brown in totana, taosana, and powelli. The dark mark anterior to the ocellus nearly always comnects to an apical dash of the same color, often connects to costa, and frequently extends along distal edge of ocellus toward tormus, forming a distinctive Y-shaped mark. The shapes of the valva (Fig. 20) and sterigma (Fig. 37) separate popana from the other species considered here.

Description. Head: Lower frons white, scales of upper frons and vertex long, gray to brownish gray medially, with lighter apices; labial palpus with basal scgment and medial surface of second segment white, lateral surface of second segment gray to brownish gray; third segment concealed by scales of second segment; antenna grayish white, often darker distally. Thorax: Dorsal surface and tegulae with brownish-gray, white-tipped scales, ventral surface white, legs with anterior surfaces brownish gray, posterior surfaces white, tarsal segments with white distal annulations. Forewing (Fig. 11): 0 FWL $8-9.8 \mathrm{~mm}$ (mean $=5.6, \mathrm{n}=12$ ), $\mathrm{AR}=3.21, \mathrm{CFR}=0.31, \%$ FWL $6.7-$ 8.3 mm (mean $=7.4, \mathrm{n}=19$ ), $\mathrm{AR}=3.21$; costa weakly convex, apex acute, termen weakly convex; dorsal surface white with brownishblack to brownish-gray markings, basal and median areas white and variably irrorated with brownish gray, an outwardly oblique subbasal mark extending from dorsum to cell, a broken median fascia consisting of three marks, the first at mid costa, the second at distal end of cell, the third semitriangular and projecting anteriorly from pretomal portion of dorsum along basal margin of ocellus, the first two median marks often connected, the latter two usually separated by narrow band of white scales, an elongate patch of white-tipped, black to brownish-gray scales anterior to ocellus, usually connected to apex by oblique dash of similar coloration; ocellus with basal, distal and tornal margins pale pinkish brown, central field a narrow, vertical, light brown streak, crossed longitudinally by ca. 4 black dashes; distal one half of costa with four paired white strigulae, costal fold of male dark gray, fringe scales white basally and apically, black to brownishgray medially. Abdomen: Females with dark gray scales on posterior margin of eighth segment. Hindwing: Uniformly brownish gray fringe lighter: Male genitalia (Fig. 20): Uncus dorsally setose and semitriangular, apex sometimes weakly indented; sociu long and setose, curving dorsally; aedeagus long, tapering distally, vesica with 36 deciduous comuti $(n=4)$; valva with costal margin concave, apes nearly right angled but rounded, distal margin convex with 2-3 stout spines, ventral angle developed into triangrlar lobe supporting 1 or 2 stout spines, ventral invagination broad and shallow; $\mathrm{NR}=0.7$, cucullus with medial surface moderately setose, sacculus sparsely. setose, margin of basal opening with weakly developed setose projection. Female genitialia (Fig. 29): Papillae anales facing ventrolaterally and densely setose, surfaces finely ridged transversely; medial margins sinuate, setae on lateral margins long, curving ventrally, those near anal opening shorter, with hooked apices: sterigna (Fig. 37) with anterior margin very weakly sclerotized, lamella postvaginalis extending laterally to width of ca. $3 \times$ ostium diameter, widening posteriorly, length ca. $0.5 \times$ width, posterior inargin with scalloped appearance due to sharply acute posterolateral comers and concave medial invagination, a ven shallow trough from mid
posterior margin to ostium. surface finely microtrichate; stemum \'11 with posterior and lateral margins strongly sclerotized, posterior margin with medial triangular projection overlapping ostium; ductus bursae of nearly uniform width, sclerotized from constriction anterior to ostium to ductus seminalis; corpus bursae with two similarly sized signa, membrane variably crinkled around signa, interior surface minutely microtrichiate.

Distribution and biology. I examined 167 specimens ( $\mathbf{1 4 6}$ of, 21 ㅇ) from the following states and counties: COLORADO: Chaffee, Grand, Larimer, Mesa, IDAHO: Blaine; MONTANA; NEVADA: Lander, White Pine; NEW MEXICO: Taos; UTAH: Cache, Garfield, San Juan, Tooele, Uintah; WYOMING: Albany, Carbon, Fremont, Park, Sublette, Teton. They document a flight period from early June to late August. No larval host has been reported.

## Pelochrista powelli Wright, new species

(Figs. 12, 22, 23, 30, 34)
Diagnosis. This moth has a pale brown appearance.
By contrast, popana is brownish-black to gray, with considerable contrast between markings and ground color. The forewing pattern of powelli lacks the chevron shaped mark at the end of the cell in totana and the orange-brown scaling along the costa and along $1 \mathrm{~A}+2 \mathrm{~A}$ in taosana. In females of powelli, the scales on the posterior margin of the eighth abdominal segment are brown and inconspicuous, they are brownish-black to gray in popana. The $v$-shaped posterior margin of the sterigma separates powelli from the other species considered here.

Description. Head: Lower frons pale tan, scales of vertex white distally, shading to tan basally; labial palpus tan to pale brown; antenna pale tan. Thorax: Dorsal surface pale brown, ventral surface pale tan, legs with anterior surfaces pale brown, posterior surfaces white to tan, distal ends of tarsal segments ringed with pale tan. Forewing (Fig. 12): ${ }^{a}$ FWL $7.1-10.5 \mathrm{~mm}($ mean $=8.7, n=10), \mathrm{AR}=3.3, \mathrm{CFR}=0.26$, क FWL 8.2-9.2 mm (mean $=8.8, \mathrm{n}=5$ ), $\mathrm{AR}=3.11$; costa weakly convex, apex acute, termen straight to weakly convex; dorsal surface pale tan with brown markings, a brownish-black subbasal mark on fold, a thin brown line along fold from subbasal mark to tornus, a narrow, triangulate, brown mark based on fold and projecting toward apex along basal margin of ocellus, an elongate patch of white-tipped brownish-black scales anterior to ocellus, extending basally throngh distal one-half of cell, usually constricted and darker at distal end of cell; ocellus obscure, variably bordered on basal, distal and tomal margins with pale pinkish-tan bars, central field white, crossed by 4-6 brownish-black dashes that are often joined in zig-zag pattem: distal two-thirds of costa with numerous white strigulae, delineated by brown costal marks and thin brown striae, male costal fold brownish black; termen with band of white-tipped brownish-black scales extending from apex to tornus, fringe scales lighter with pale-brown medial markings. Hindwing: Pale gray-brown with paler fringe. Male genitalia (Figs. 22, 23): Uncus semitriangular with rounded apex, dorsal surface setose; tegumen long, dorsolateral shoulders well developed and hunched; socii long, flat, tapering distally, and moderately setose; gnathos a narrow band; aedcagus long, tapered distally, vesica with $4-9$ deciduous cornuti ( $\mathrm{n}=7$ ), valva with costal margin weakly concave, apex rounded to angular, distal margin convex, ventral angle with triangular projection supporting stout spine, neck long and narrow, $\mathrm{NR}=0.5$, ventral invagination broad and moderate, cucullus with densely setose medial surface and 3-5 stout spines on distal margin, sacculus moderately setose, margin of basal


Fig. 30. Female genitalia. 30, P. powelli, slide DJW 1024.
opening with weakly developed setose projection. Female genitalia (Fig. 30): Papillae anales facing ventrolaterally and densely setose, medial margins sinuate, surfaces very finely ridged transversely, setae on lateral margins long and curving ventrally, those near anal opening shorter with hooked apices; posterior one-half of tergum VIII with 45 rows of setae; sterigma (Fig.34) with anterior margin very weakly sclerotized, lamella postvaginalis developed posterolaterally into triangular projections, posterior margin with $v$-shaped medial invagination, surface finely microtrichiate; stermum VII with posterior margin concavely invaginated except for mild, convex, medial, projection overlapping ostium; ductus bursae narrow, constricted anterior to ostium; corpus bursae with two signa, inner surface minutely microtrichiate.

Holotype. ${ }^{\text {By }}$, Idaho, Oneida Co., Curlew NC, 4 mi. ENE of Holbrook, Jctn. Forest Rds. 056 and 057, 5050', 7 July 2001, D. J. Wright, genitalia slide DJW 1032, deposited in USNM. Type locality at $42^{\circ} 11.35^{\prime} \mathrm{N}, 112^{\circ} 34.92^{\prime} \mathrm{W}$.

Paratypes. IDAHO: Oneida Co., Curlew NG, 4 mi . ENE of Holbrook, $5050^{\prime}$, D. J. Wright, 25 July 2003 (2 d, genitalia slide DJV1027), 26 July 2003 (2 $\delta, 1$ ) ); Oneida Co., Curlew NG, T14S R32E S30, D. J. Wright, 28 July 2003 ( 4 \& , genitalia slides DJW1024, 1029). UTAH: Daggett Co., 4 mi. S. of Manila, C. J. Balogh, 20 July


FIGS. 31-37. Sterigmata of specimens illustrated in Figs. 24-30. 31, E. nordini. 32, E. totana. 33, P. emaciatana. 34, P. pouelli. 35, E. piperata. 36, E. larana. 37, P. popana.

1994 (503, genitalia slides DJV 697, 705, 707); Juab Co., Eureka, Tom Spalding, 18 August 1911 ( 10 , genitalia slide DJVI069); Garfield Co, 3 mi . W. Bryce Jct., $2300 \mathrm{~m}, \mathrm{~J}$. A. Powell, 2S/29 June 1992 (1 dै, genitalia slide EME 5755); Garfield Co., Kings Cr. campgr., 15 km SW Bryce Jct., $2300 \mathrm{~m}, \mathrm{~J}$. A. Powell, IS July 1993 ( 12 o $^{\circ}$, genitalia slide EME 5756). Paratype depositories: BMNH, CNC, EME, GJB, USNM, DJW.

Etymology. This species is named after J. A. Powell, who collected nearly half of the specimens in the type series
Distribution and biology. The 29 specimens ( $244^{6}$, 5 ) ${ }^{\text {) }}$ reported above were collected in southeastern Idaho and Utah, suggesting a Great Basin distribution for this insect. The type locality is open sage brush habitat. The larval host is unknown.

Comments. The shape of the male valva is variable (Figs. 22, 23), the cucullus illustrated in Fig. 22 being the most angular of the nine I examined. Forewing color also varies from very light $\tan$ in the specimens from Idaho to a pale brown in those from Utah.

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